

Total soluble Neuropilin-1 ELISA

for the quantitative determination of human
soluble Neuropilin-1 in serum, EDTA plasma, heparin plasma, and citrate plasma
Cat. No. BI-20409. 12 x 8 tests

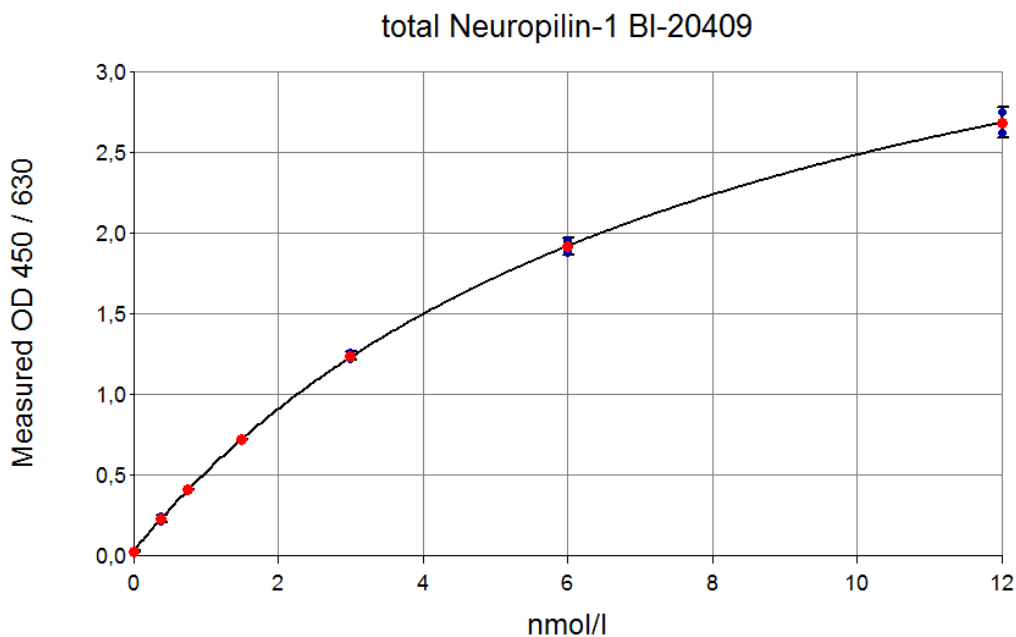
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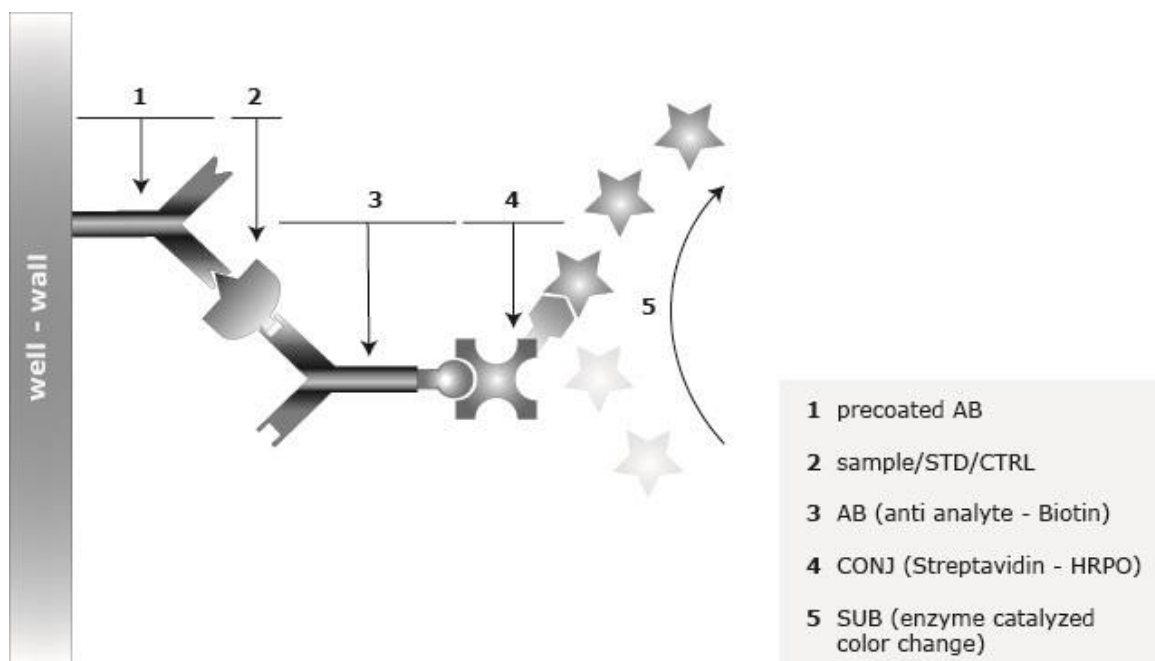
ASSAY CHARACTERISTICS Summary

| | | | |
|---|--|--------------------------|------------------------|
| Method | Sandwich ELISA, HRP/TMB, 12x8-well strips | | |
| Sample type | Serum, EDTA plasma, heparin plasma, and citrate plasma | | |
| Standard range | 0 to 12 nmol/l (0 / 0.37 / 0.75 / 1.5 / 3 / 6 / 12 nmol/l) 7 standards and 2 controls in a serum matrix. | | |
| Conversion factor | soluble Neuropilin-1: 1 ng/ml = 0.014 nmol/l; 1 nmol/l=69.7 ng/ml (MW: 69.7 kDa) | | |
| Sample volume | 10 µl / well | | |
| Incubation time, temp. | 30 min / 2 h / 1 h / 30 min, room temperature | | |
| Sensitivity | LOD (0 pmol/l + 3 SD): 0.09 nmol/l; LLOQ: 0.09 nmol/l | | |
| Specificity | <p>The assay is optimized to detect soluble total Neuropilin-1 (NRP1) in human plasma and serum. This assay recognizes endogenous and recombinant human soluble Neuropilin-1 (isoform 2 and 3). The total soluble Neuropilin-1 ELISA utilizes a monoclonal anti-human Neuropilin-1 antibody that binds to a linear epitope close to the N-terminus in the CUB 1 domain of the Neuropilin-1 molecule. The polyclonal detection antibody binds to multiple linear epitopes, distributed over the entire Neuropilin-1 molecule.</p> <p>The human sequence homology between NRP1 and NRP2 is low. The amino acid sequence in the respective binding regions of the antibodies show no homology with NRP2. Thus a cross reactivity to NRP2 is not expected.</p> | | |
| Precision | Intra-assay (n=6) ≤ 11%, Inter-assay (n=12) ≤ 10% | | |
| Spike/Recovery (recombinant 1.5 + 6 nmol/l Neuropilin-1) | <u>Average % recovery</u> | <u>1.5 nmol/l</u> | <u>6 nmol/l</u> |
| | Serum (n=6) | 90 | 92 |
| | EDTA plasma (n=6) | 91 | 93 |
| | Citrate plasma (n=1) | 98 | 108 |
| | Heparin plasma (n=1) | 115 | 97 |
| Dilution linearity of endogenous soluble Neuropilin-1 | <u>Average % of expected of dilution:</u> | <u>1+1</u> | <u>1+3</u> |
| | Serum (n=6): | 95 | 104 |
| | EDTA plasma (n=6): | 107 | 110 |
| | Citrate plasma (n=1): | 101 | 102 |
| | Heparin plasma (n=1): | 99 | 104 |
| Values of apparently healthy individuals | <p>Median serum (n=24) = 2.0 nmol/l Median EDTA plasma (n=24) = 1.7 nmol/l Median heparin plasma (n=24) = 1.9 nmol/l Median citrate plasma (n=24) = 1.7 nmol/l</p> <p>Each laboratory should establish its own reference range for the samples under investigation. Do not change sample type during the study.</p> | | |

TYPICAL STANDARD CURVE



PRINCIPLE OF THE ASSAY



CAB Coating antibody: Polyclonal anti-human Neuropilin-1 IgG
 DAB Detection antibody: Monoclonal mouse antibody IgG, biotin labelled
 AG Antigen: recombinant human Neuropilin-1 protein (AA22-AA644 of Uniprot ID O14786-2, isoform 2)

INFORMATION on the ANALYTE

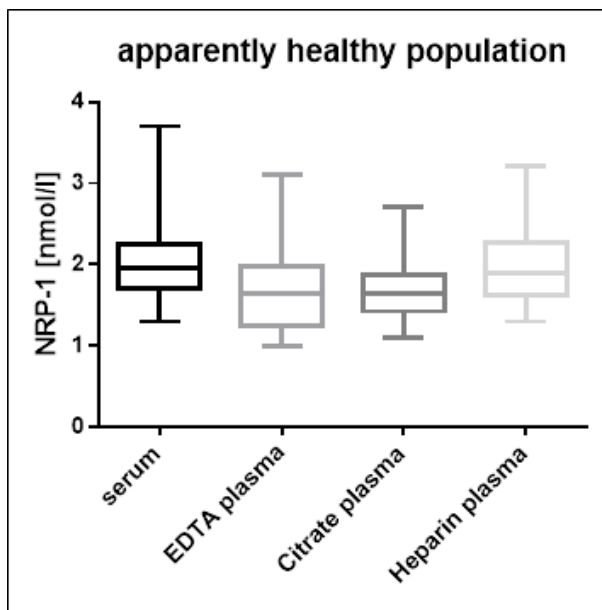
Neuropilin-1 (NRP1) is a single-pass transmembrane glycoprotein of 923 amino acids, composed of a large extracellular region, a short transmembrane domain and a short cytoplasmic tail <https://www.uniprot.org/uniprot/O14786>. Due to alternative splicing or shedding, the extracellular region can be released into circulation as soluble Neuropilin. NRP1 is an essential cell surface receptor functioning in many key biological processes including the cardiovascular, neuronal, and immune systems (1,2). Multiple ligands bind to the extracellular region of NRP1, like class III semaphorins which have a key role in axonal guidance, or members of the VEGF family of angiogenic cytokines. Ligand-binding to transmembrane NRP1, which has co-receptor function, leads to signaling via receptor proteins containing a PDZ domain. In contrast, ligand-binding to soluble Neuropilin-1 (sNRP1) has antagonistic properties by acting as decoy (1,3). NRP1 is expressed by a variety of cells and tissues. For instance, the transmembrane protein is expressed by neuronal cells, endothelial cells, vascular smooth muscle cells, cardiomyocytes, multiple tumor cell lines and neoplasms, osteoblasts, naïve T cells or platelets. Expression of soluble Neuropilin-1 is further described in a variety of non-endothelial cells, e.g. in liver hepatocytes and kidney distal and proximal tubules. NRP1 is implicated in a multitude of physiological and pathological settings, e.g. in axon guidance, vascularization, tumor growth or regeneration and repair (4-9). Neuropilin-1 is described to stimulate osteoblast differentiation, to act as potential biomarker for the prediction of heart failure outcome or to play a role in renal fibrogenesis (6, 10,11). As a co-receptor for VEGF, NRP1 is a potential target for cancer therapies (12). The Neuropilin-1 enzyme immunoassay is a four hour ELISA to quantify human total soluble Neuropilin-1 (sNRP1). The assay is validated for human serum and plasma samples (EDTA, citrate, heparin) (13) (see validation data: www.bmgrp.com). To remove potentially bound ligands, samples are pre-treated with guanidine hydrochloride before testing. Recombinant human soluble Neuropilin-1, isoform 2, is used as calibrator.

SAMPLE VALUES

soluble Neuropilin-1 values in an apparently healthy cohort

| | Neuropilin-1 [nmol/l] | | | |
|----------------|-----------------------|-----------------------|--------------------------|--------------------------|
| | Serum (n=24) | EDTA plasma (n=24) | Citrate plasma (n=24) | Heparin plasma (n=24) |
| Mean | 2.0 | 1.7 | 1.7 | 2.0 |
| Median | 2.0 | 1.7 | 1.7 | 1.9 |
| 5% Percentile | 1.3 | 1.0 | 1.1 | 1.3 |
| 95% Percentile | 3.4 | 3.0 | 2.6 | 3.1 |
| Minimum | 1.3 | 1.0 | 1.1 | 1.3 |
| Maximum | 3.7 | 3.1 | 2.7 | 3.2 |

It is recommended to establish the normal range for each laboratory.

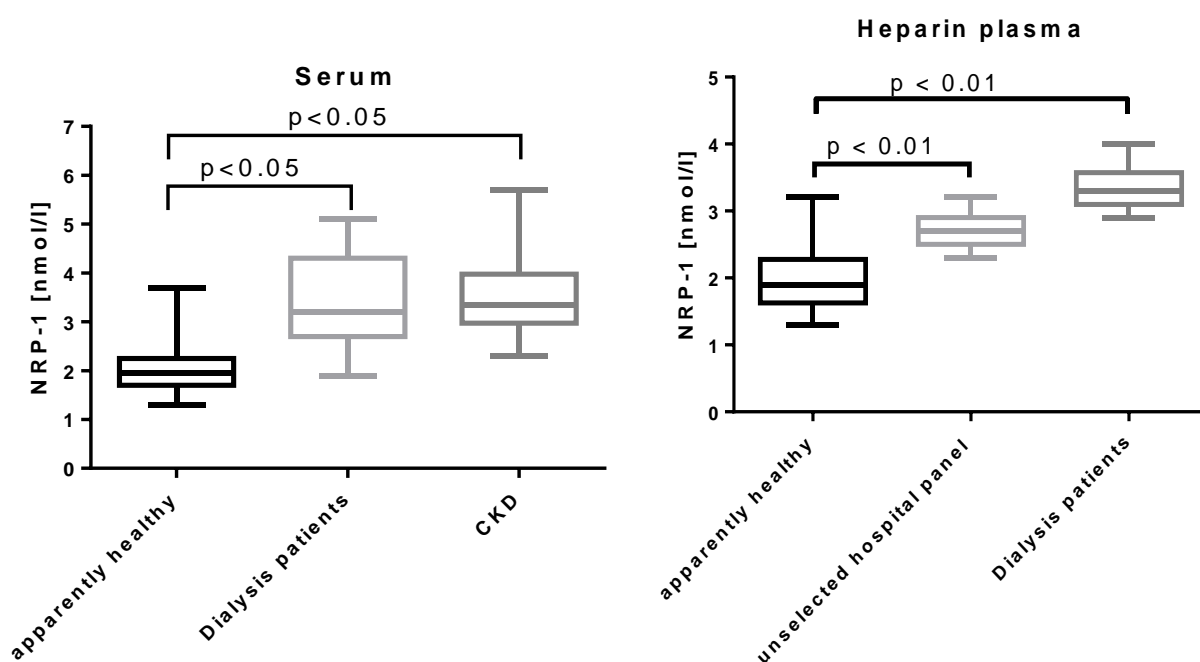


soluble Neuropilin-1 values in **serum** samples

| | Neuropilin-1 [nmol/l] | | |
|----------------|---------------------------|--------------------------|------------|
| | Apparently healthy (n=24) | Dialysis patients (n=15) | CKD (n=22) |
| Mean | 2.0 | 3.4 | 3.5 |
| Median | 2.0 | 3.2 | 3.4 |
| 5% Percentile | 1.3 | 1.9 | 2.3 |
| 95% Percentile | 3.4 | 5.1 | 5.6 |
| Minimum | 1.3 | 1.9 | 2.3 |
| Maximum | 3.7 | 5.1 | 5.7 |

soluble Neuropilin-1 values in **heparin plasma** samples

| | Neuropilin-1 [nmol/l] | | |
|----------------|---------------------------|---------------------------------|--------------------------|
| | Apparently healthy (n=24) | Unselected hospital panel (n=8) | Dialysis patients (n=16) |
| Mean | 2.0 | 2.7 | 3.3 |
| Median | 1.9 | 2.7 | 3.3 |
| 5% Percentile | 1.3 | 2.3 | 2.9 |
| 95% Percentile | 3.1 | 3.2 | 4 |
| Minimum | 1.3 | 2.3 | 2.9 |
| Maximum | 3.2 | 3.2 | 4 |



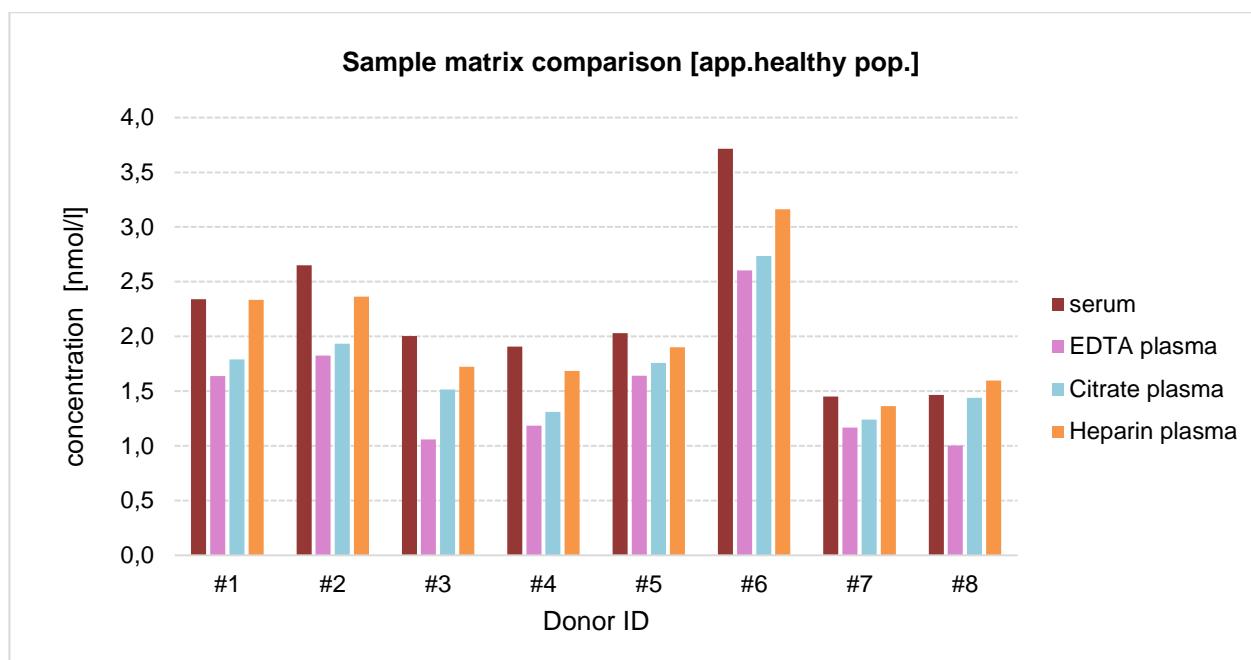
MATRIX COMPARISON

Comparison of Neuropilin-1 serum and plasma sample values from apparently healthy individuals

8 samples of apparently healthy individuals were prepared, each sample derived from one donor. Samples were assayed and the concentrations of the samples were compared.

| Donor ID | Neuropilin-1 [nmol/l] | | | | CV [%] | |
|--------------------|-----------------------|-------------|----------------|----------------|--------------|-------------|
| | Serum | EDTA plasma | Citrate plasma | Heparin plasma | all matrices | only plasma |
| #1 | 2.3 | 1.6 | 1.8 | 2.3 | 16 | 16 |
| #2 | 2.6 | 1.8 | 1.9 | 2.4 | 15 | 11 |
| #3 | 2.0 | 1.1 | 1.5 | 1.7 | 22 | 19 |
| #4 | 1.9 | 1.2 | 1.3 | 1.7 | 19 | 15 |
| #5 | 2.0 | 1.6 | 1.8 | 1.9 | 8 | 6 |
| #6 | 3.7 | 2.6 | 2.7 | 3.2 | 14 | 8 |
| #7 | 1.5 | 1.2 | 1.2 | 1.4 | 8 | 6 |
| #8 | 1.5 | 1.0 | 1.4 | 1.6 | 16 | 19 |
| Mean CV [%] | | | | | 15 | 13 |

Graph showing soluble Neuropilin-1 levels in various sample matrices



ASSAY PERFORMANCE CHARACTERISTICS

RECOVERY

Summary of data showing mean recovery of soluble Neuropilin-1

| Matrix | Mean S/R [%] | |
|----------------------|--------------|------------|
| | + 1.5 nmol/l | + 6 nmol/l |
| Serum (n=6) | 90 | 92 |
| EDTA plasma (n=6) | 91 | 93 |
| Citrate plasma (n=1) | 98 | 108 |
| Heparin plasma (n=1) | 115 | 97 |

Experiments:

Recovery of spiked samples was tested by adding 2 concentrations of human recombinant Neuropilin-1 (1.5 + 6 nmol/l) to different human serum and plasma sample matrices.

Data showing spike/recovery of human **serum** samples

| Sample ID | Spike Neuropilin-1 [nmol/l] | | | S/R [%] | |
|-----------|-----------------------------|-----|-------------------|-----------|-----------|
| | 0 | 1.5 | 6 | 1.5 | 6 |
| #S1 | 1.5 | 2.1 | 6.1 | 91 | 90 |
| #S2 | 2.0 | 2.4 | 6.8 | 94 | 97 |
| #S3 | 2.6 | 3.0 | 7.8 | 113 | 109 |
| #S4 | 1.6 | 2.1 | 6.3 | 85 | 92 |
| #S5 | 1.8 | 2.2 | 5.8 | 84 | 81 |
| #S6 | 2.6 | 2.4 | 6.3 | 75 | 84 |
| | | | Mean R [%] | 90 | 92 |

Data showing spike/recovery of human **EDTA plasma** samples

| Sample ID | Spike Neuropilin-1 [nmol/l] | | | S/R [%] | |
|-------------------|-----------------------------|-----|-----|-----------|-----------|
| | 0 | 1.5 | 6 | 1.5 | 6 |
| #E1 | 1.4 | 2.1 | 6.6 | 93 | 98 |
| #E2 | 1.8 | 2.4 | 7.1 | 98 | 103 |
| #E3 | 2.1 | 2.6 | 7.6 | 107 | 109 |
| #E4 | 1.5 | 2.2 | 5.7 | 95 | 82 |
| #E5 | 1.6 | 2.1 | 5.7 | 90 | 83 |
| #E6 | 2.2 | 2.4 | 6.2 | 88 | 85 |
| Mean R [%] | | | | 95 | 93 |

Data showing spike/recovery of human **citrate plasma** sample

| Sample ID | Spike Neuropilin-1 [nmol/l] | | | S/R [%] | |
|-----------|-----------------------------|-----|-----|---------|-----|
| | 0 | 1.5 | 6 | 1.5 | 6 |
| #C1 | 1.9 | 2.4 | 7.4 | 98 | 108 |

Data showing spike/recovery of human **heparin plasma** sample

| Sample ID | Spike Neuropilin-1 [nmol/l] | | | S/R [%] | |
|-----------|-----------------------------|-----|-----|---------|----|
| | 0 | 1.5 | 6 | 1.5 | 6 |
| #H1 | 2.2 | 2.8 | 6.9 | 115 | 97 |

LINEARITY

Dilution linearity of samples containing endogenous soluble Neuropilin-1

| Matrix | Mean R of dilution steps [%] | | |
|----------------------|------------------------------|-----|-----|
| | 1+1 | 1+3 | 1+7 |
| Serum (n=6) | 95 | 104 | 114 |
| EDTA plasma (n=6) | 107 | 110 | 115 |
| Citrate plasma (n=1) | 101 | 102 | 111 |
| Heparin plasma (n=1) | 99 | 104 | 110 |

- ▶ We recommend diluting high measuring samples (outside of the calibration range) with STD1 (0 nmol/l, supplied in the kit).

Experiment:

Dilution linearity was assessed by serially diluting samples containing endogenous soluble Neuropilin-1 with STD1.

Data showing the dilution of endogenous Neuropilin-1 in **serum** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-------------------|-----------------------|-----|-----|-----|-----------|------------|------------|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #S1 | 2.7 | 1.6 | 0.9 | 0.5 | 122 | 129 | 148 |
| #S2 | 5.1 | 2.7 | 1.5 | 0.8 | 107 | 115 | 123 |
| #S3 | 4.5 | 1.8 | 1.1 | 0.6 | 80 | 95 | 99 |
| #S4 | 5.0 | 2.4 | 1.3 | 0.7 | 95 | 99 | 107 |
| #S5 | 5.4 | 2.2 | 1.2 | 0.7 | 82 | 90 | 104 |
| #S6 | 4.0 | 1.7 | 1.0 | 0.5 | 86 | 97 | 104 |
| Mean R [%] | | | | | 95 | 104 | 114 |

Data showing the dilution of endogenous Neuropilin-1 in **EDTA plasma** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-------------------|-----------------------|-----|-----|-----|------------|------------|------------|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #E1 | 2.8 | 1.6 | 0.9 | 0.5 | 117 | 127 | 137 |
| #E2 | 2.7 | 1.3 | 0.7 | 0.4 | 98 | 105 | 106 |
| #E3 | 3.4 | 1.9 | 0.9 | 0.5 | 110 | 111 | 114 |
| #E4 | 3.2 | 1.7 | 0.9 | 0.4 | 108 | 114 | 111 |
| #E5 | 3.3 | 1.8 | 0.9 | 0.5 | 111 | 107 | 117 |
| #E6 | 3.9 | 2.0 | 0.9 | 0.5 | 101 | 97 | 105 |
| Mean R [%] | | | | | 107 | 110 | 115 |

Data showing the dilution of endogenous Neuropilin-1 in **citrate plasma** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-----------|-----------------------|-----|-----|-----|-------|-----|-----|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #C1 | 2.7 | 1.4 | 0.7 | 0.4 | 101 | 102 | 111 |

Data showing the dilution of endogenous Neuropilin-1 in **heparin plasma** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-----------|-----------------------|-----|-----|-----|-------|-----|-----|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #H1 | 3.1 | 1.5 | 0.8 | 0.4 | 99 | 104 | 110 |

Dilution linearity of samples containing recombinant soluble Neuropilin-1

| Matrix | Mean R of dilution steps [%] | | |
|----------------------|------------------------------|-----|-----|
| | 1+1 | 1+3 | 1+7 |
| Serum (n=3) | 95 | 98 | 99 |
| EDTA plasma (n=3) | 84 | 96 | 94 |
| Citrate plasma (n=1) | 77 | 90 | 88 |
| Heparin plasma (n=1) | 95 | 104 | 104 |

Recommendations for sample dilution:

- ▶ We recommend diluting high measuring samples (outside of the calibration range) with STD1 (0 nmol/l, supplied in the kit).

Experiment:

Dilution linearity was assessed by serially diluting samples containing recombinant soluble Neuropilin-1 with STD1.

Data showing the dilution of recombinant Neuropilin-1 in **serum** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-------------------|-----------------------|-----|-----|-----|-----------|-----------|-----------|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #S1 | 6.1 | 3.3 | 1.6 | 0.8 | 108 | 102 | 110 |
| #S2 | 6.8 | 3.4 | 1.6 | 0.8 | 99 | 92 | 90 |
| #S3 | 7.8 | 3.1 | 2.0 | 0.9 | 79 | 100 | 97 |
| Mean R [%] | | | | | 95 | 98 | 99 |

Data showing the dilution of recombinant Neuropilin-1 in **EDTA plasma** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-------------------|-----------------------|-----|-----|-----|-----------|-----------|-----------|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #E1 | 6.6 | 2.9 | 1.7 | 0.8 | 87 | 101 | 101 |
| #E2 | 7.1 | 3.1 | 1.7 | 0.8 | 86 | 96 | 89 |
| #E3 | 7.6 | 3.0 | 1.7 | 0.9 | 79 | 92 | 91 |
| Mean R [%] | | | | | 84 | 96 | 94 |

Data showing the dilution of recombinant Neuropilin-1 in **citrate plasma** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-----------|-----------------------|-----|-----|-----|-------|-----|-----|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #C1 | 7.4 | 2.9 | 1.7 | 0.8 | 77 | 90 | 88 |

Data showing the dilution of recombinant Neuropilin-1 in **heparin plasma** samples

| Sample ID | Neuropilin-1 [nmol/l] | | | | R [%] | | |
|-----------|-----------------------|-----|-----|-----|-------|-----|-----|
| | ref | 1+1 | 1+3 | 1+7 | 1+1 | 1+3 | 1+7 |
| #H1 | 6.9 | 3.3 | 1.8 | 0.9 | 95 | 104 | 104 |

PRECISION

Intra-assay precision & Inter-assay precision

Intra-assay (n=6) ≤ 11%, Inter-assay (n=) ≤ 10%

Intra-assay: 2 samples of known concentrations were tested 6 times with 1 kit lot by 1 operator.
Inter-assay: 2 samples of known concentrations were tested 12 times with 2 different kit lots on 3 days by 3 different operators.

| Intra-assay (n=6) | Sample 1 | Sample 2 | Inter-assay (n=12) | Sample 1 | Sample 2 |
|--------------------------|-----------------|-----------------|---------------------------|-----------------|-----------------|
| Mean (nmol/l) | 0.8 | 6.2 | Mean (pmol/l) | 0.8 | 6.2 |
| SD (nmol/l) | 0.1 | 0.4 | SD (pmol/l) | 0.08 | 0.36 |
| CV (%) | 11 | 7 | CV (%) | 10 | 6 |

SENSITIVITY

Limit of detection (LOD)

The LOD is defined as the mean value of the back calculated concentration plus three times the standard deviation. The LOD of the total soluble Neuropilin-1 ELISA is **0.09 nmol/l**.

Lower limit of quantification (LLOQ)

The lower limit of quantification is defined as the accuracy of the back calculated concentrations and shall not exceed $\pm 25\%$ (acc. to ICH [Ref. 1]).

The LLOQ of the total soluble Neuropilin-1 ELISA is **0.09 nmol/l**.

SAMPLE STABILITY

Sample preparation

Collect venous blood samples by using standardized blood collection tubes. Perform serum and plasma separation by centrifugation according to supplier's instructions of the blood collection devices as soon as possible. The acquired serum and plasma samples should be measured as soon as possible. For longer storage aliquot samples and store at -25°C or lower for long time storage at -80°C . All samples should undergo only 5 freeze-thaw cycles.

Freeze/thaw of plasma samples containing endogenous soluble Neuropilin-1

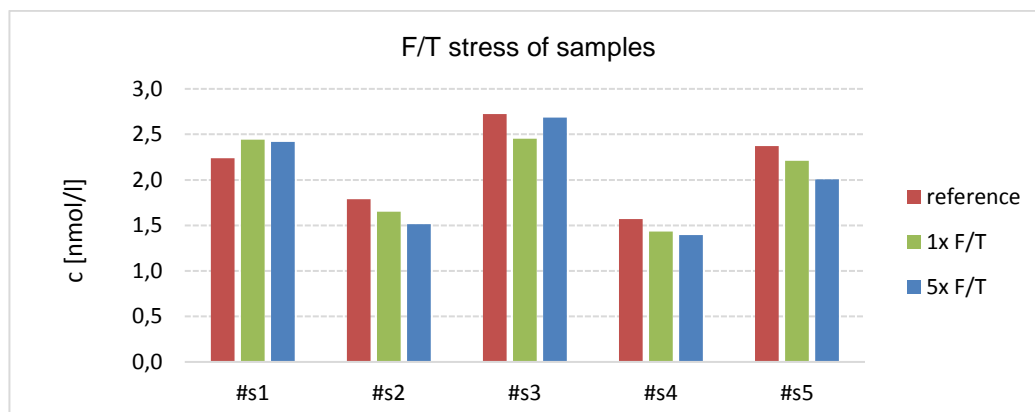
Five serum samples were aliquoted and freeze-thaw stressed. The reference samples are freeze-thawed once. Samples can undergo 5 freeze-thaw cycles. The mean recovery of sample concentrations stressed by 5 F/T cycles is 95%.

Samples can undergo 5 freeze-thaw cycles.

Data showing Neuropilin-1 concentrations of samples after freeze-thaw cycles:

| Sample ID | Neuropilin-1 [nmol/l] | | | R [%] 5 F/T vs ref |
|-----------|-----------------------|-----|-------------------|-----------------------|
| | Reference | 1x | 5x | |
| #S1 | 2.2 | 2.4 | 2.4 | 108 |
| #S2 | 1.8 | 1.7 | 1.5 | 85 |
| #S3 | 2.7 | 2.5 | 2.7 | 99 |
| #S4 | 1.6 | 1.4 | 1.4 | 89 |
| #S5 | 2.4 | 2.2 | 2.0 | 85 |
| | | | Mean R [%] | 93 |

Graph showing F/T cycles of serum samples



CHARACTERIZATION OF THE ANTIBODIES

The total soluble Neuropilin-1 ELISA utilizes a monoclonal anti-human Neuropilin-1 antibody that binds to a linear epitope close to the N-terminus in the CUB 1 domain of the Neuropilin-1 molecule. The polyclonal detection antibody binds to multiple linear epitopes, distributed over the entire Neuropilin-1 molecule.

DAB Detection Antibody: Monoclonal mouse anti-Neuropilin-1 antibody-HRP binds to a linear epitope close to the N-terminus in the CUB 1 domain of the Neuropilin-1 molecule.

CAB Coating Antibody: Polyclonal sheep anti-human Neuropilin-1 antibody binding to multiple linear epitopes distributed over the entire Neuropilin-1 molecule (between AA81-AA630).

SPECIFICITY

The assay is optimized to detect soluble total Neuropilin-1 (NRP1) in human plasma and serum. The specific interaction of the analyte with the coating and the detection antibody was analysed. This assay recognizes endogenous (natural) and recombinant human soluble Neuropilin-1 (isoform 2 and 3).

The human sequence homology between NRP1 and NRP2 is low. The amino acid sequence in the respective binding regions of the antibodies show no homology with NRP2. Thus a cross reactivity to NRP2 is not expected.

ISOFORMS

Isoform1 is not soluble in blood.

Isoforms 2 (longer) and 3 (shorter, C-terminal domain missing), which are generated by alternative splicing, are both soluble and detected by this assay (cat. no. BI-20409).

CALIBRATION

This immunoassay is calibrated against recombinant human soluble Neuropilin-1 protein (AA22-AA644 of O14786-2 (Uniprot ID)).

VALIDATION GUIDELINES

The assay is fully validated for human serum and plasma samples according to ICH Q2 (R1) (8).

LITERATURE

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Available on our Website www.bmgrp.com

[Instructions for use \(IFU\)](#)

Protocol for urine samples
Protocol for cell culture supernatants samples
Protocol for non-human samples
Material Safety Data Sheet (MSDS)
Biomedica Neuropilin-1 ELISA – Info Leaflet

Date: May 2018

MEASUREMENT OF NEUROPILIN-1 in human URINE, CELL CULTURE SUPERNATANTS and NON-HUMAN SAMPLES

The following experiments have been performed to test the use of the Neuropilin-1 assay (cat. no. BI-20409) in human urine samples, human cell culture supernatants and non-human samples.

1. MEASUREMENT of NEUROPILIN-1 in HUMAN URINE SAMPLES

Note: the experiments performed to measure total soluble Neuropilin-1 in urine samples are not a full validation but are merely a performance check.

Performance check:

Urine samples (n=6) from apparently healthy subjects and from patients with kidney disease were assayed with the total soluble Neuropilin-1 ELISA following the standard protocol (including GuHCl treatment) using undiluted urine. Sample concentrations were calculated by using STD1-STD7.

- Endogenous Neuropilin-1 was undetectable in tested samples (cohorts from apparently healthy and kidney disease) thus no test on specificity (competition) was carried out.
- Urine samples can be spiked with recombinant human Neuropilin-1. The average recovery of 6 human urine samples is 105%.
- Urine samples spiked with recombinant human Neuropilin-1 can be diluted 1+1 with STD1 (0 nmol/l) that is supplied in the kit. The average recovery of 6 human urine samples is 94%.

RECOVERY

Recovery was assessed by adding STD7 (supplied in the kit, final concentration 6 nmol/l human recombinant Neuropilin-1) directly to 6 different human urine samples (ratio 1+1).

Data showing spike/recovery of human urine samples:

| Sample ID | Neuropilin-1 [nmol/l] | | S/R [%] |
|------------------|------------------------------|-------------------|----------------|
| | Reference | + 6 nmol/l | |
| #U1 | 0.1 | 6.1 | 100 |
| #U2 | 0 | 6.3 | 105 |
| #U3 | 0 | 6.5 | 108 |
| #U4 | 0 | 6.3 | 105 |
| #U5 | 0 | 6.5 | 108 |
| #U6 | 0 | 6.3 | 105 |
| | | Mean R [%] | 105 |

LINEARITY

Dilution linearity was assessed by diluting urine samples spiked with 6 nmol/l recombinant Neuropilin-1 with STD1 (0 nmol/l, supplied in the kit).

Data showing the dilution of recombinant Neuropilin-1 in urine samples:

| Sample ID | Neuropilin-1 [nmol/l] | | Dil R [%] |
|-----------|-----------------------|---------|-----------|
| | Reference | Dil 1+1 | |
| #U1 | 6.1 | 2.6 | 87 |
| #U2 | 6.3 | 2.8 | 89 |
| #U3 | 6.5 | 3.2 | 98 |
| #U4 | 6.3 | 3.0 | 96 |
| #U5 | 6.5 | 3.2 | 100 |
| #U6 | 6.3 | 3.0 | 94 |
| | Mean R [%] | | 94 |

Suggested protocol for the measurement of human Neuropilin-1 in urine samples

Follow standard protocol as indicated in the package insert:

In pre-dilution plate:

*Use **10 µl urine sample for GuHCl treatment.***

Continue with protocol as indicated in the instructions for use.

In pre-coated plate:

*Transfer **50 µl pre-treated urine sample** from pre-dilution plate into respective wells.
Swirl gently.*

*For the transfer into the coated plate it is recommended to use a multichannel pipette.
Transfer should be performed as soon as possible.*

Continue with protocol as indicated in the instructions for use.

If required, dilute samples 1+1 with STD1 (provided in the kit) prior to GuHCl treatment.

2. MEASUREMENT of NEUROFILIN-1 in CELL CULTURE SUPERNATANTS

Note: the experiments performed to measure total soluble Neuropilin-1 in cell culture supernatants are not a full validation but are merely a performance check.

Performance Check:

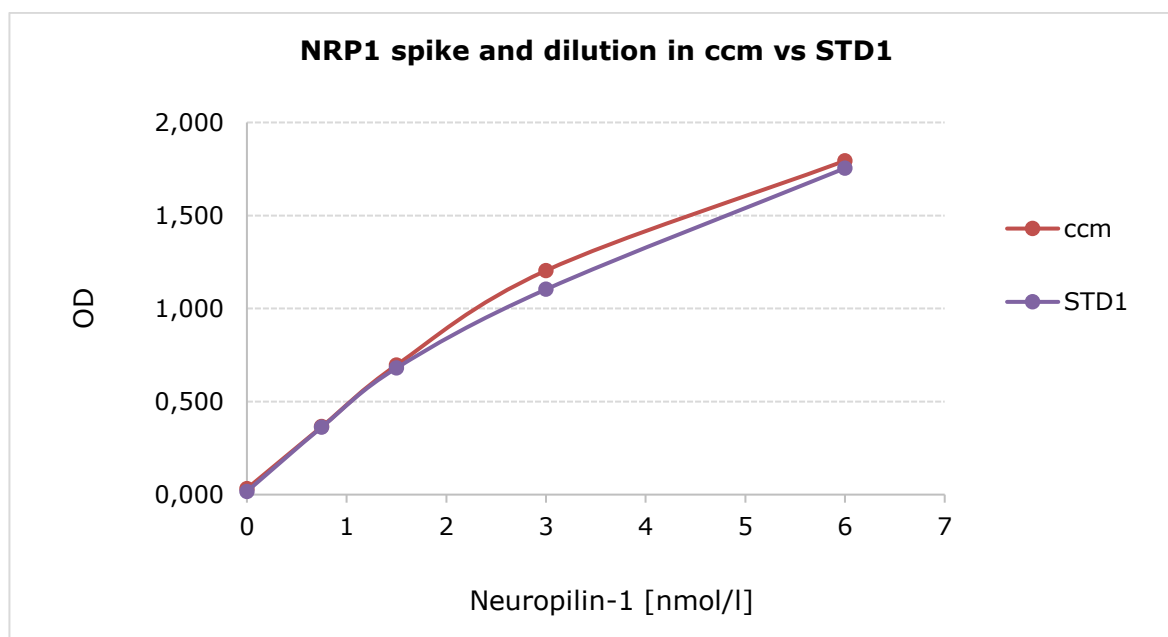
Cell culture medium (ccm: RPMI1640 containing 10% fetal calf serum) was tested undiluted and spiked by adding STD7 (provided in the kit, final concentration 6 nmol/l human recombinant Neuropilin-1) directly to cell culture medium (ratio 1+1). The spiked solution was diluted 1+1, 1+3 and 1+7 with the cell culture medium.

As a comparison, the spike recovery and dilution linearity of the standard matrix (=STD1, containing 0 nmol/l human recombinant Neuropilin-1) and the dilutions with cell culture medium is shown.

OD values of spiked and diluted cell culture medium sample and standard matrix (STD1):

| Dilution medium | Sample ID | Neuropilin-1 [nmol/l] | | | | |
|-----------------|-----------|-----------------------|------------|-----|-----|-----|
| | | Reference | + 6 nmol/l | 1+1 | 1+3 | 1+7 |
| ccm | ccm | 0 | 6.0 | 3.2 | 1.6 | 0.8 |
| ccm | STD1 | 0 | 5.8 | 2.8 | 1.5 | 0.7 |

Graph showing dilution of cell culture medium (ccm) and a comparison to the standard (STD1, containing 0 nmol/l human NRP1). Both ccm and STD1 were spiked with the same amount of human Neuropilin-1 (6 nmol/l).



Protocol for the measurement of human Neuropilin-1 in cell culture supernatants

Preparation of a cell culture medium (ccm) based standard curve:

Reconstitute STD7 in 200 µl deionized water. Leave at room temperature (18-26°C) for 15 min and mix well prior to making dilutions.

Use polypropylene tubes.

For the preparation of the cell culture medium based standards *always* use the identical cell culture medium in which the samples are based on.

- Mark tubes e.g. CC STD6, CC STD 5 ... CC STD1.
- Prepare a two-fold serial dilution to obtain STD6 to STD2.

e.g.:

Dispense 100 µl cell culture medium into vials labelled with CC STD6 to CC STD1.

Pipette 100 µl of STD 7 into tube marked as CC STD6. Mix thoroughly.

Transfer 100 µl of CC STD6 into vial marked as CC STD5. Mix thoroughly. Continue in the same fashion to obtain CC STD4 to CC STD2.

- Cell culture medium serves as the zero standard (=CC STD1, 0 nmol/l).

NOTE: after preparation of the cell culture medium (ccm) based standard curve, proceed with the protocol as indicated in the instructions for use. ALL samples including the ccm based standards must be treated with GuHCl before assaying.

Attention: Concentrations defined for CTRL A and B are only valid for measuring total soluble Neuropilin-1 in human serum or plasma. The controls cannot be used for cell culture measurements.

Suggested protocol for the measurement of human Neuropilin-1 in cell culture supernatants (cc)

Follow standard protocol as indicated in the package insert:

In pre-dilution plate:

*Use **10 µl cc sample for GuHCl treatment.***

In pre-coated plate:

*Transfer **50 µl pre-treated cc sample** from pre-dilution plate into respective wells.*

Swirl gently.

For the transfer into the coated plate it is recommended to use a multichannel pipette.

Transfer should be performed as soon as possible.

If required, dilute samples 1+1 with ccm (cell culture medium) prior to GuHCl treatment.

3. MEASUREMENT of total soluble NEUROFILIN-1 in NON-HUMAN SAMPLES

Note: the experiments performed to measure total soluble Neuropilin-1 in non-human samples are not a full validation but are merely a performance check.

The sequence of Neuropilin-1 in mammals is highly conserved.

The listed species below show a homology of >90% with human Neuropilin-1:

- Macaca mulatta (Rhesus macaque)
- Pan paniscus (Pygmy chimpanzee) (Bonobo)
- Pongo abelii (Sumatran orangutan) (Pongo pygmaeus abelii)
- Sus scrofa (Pig)
- Chlorocebus sabaeus (Green monkey) (Cercopithecus sabaeus)
- Aotus nancymae (Ma's night monkey)
- Gorilla gorilla gorilla (Western lowland gorilla)
- Neovison vison (American mink) (Mustela vison)
- Callithrix jacchus (White-tufted-ear marmoset)
- And more

Table: Sequence comparison of NRP-1 protein in mouse, human, and rat

| | | | | |
|--------|------------|-----|--|-----|
| P97333 | NRP1_MOUSE | 1 | MERGLPLLCLALALALAGAFRSDKCGGTIKIENPGYLTPGYPHSHYHSEKCEWLIQA | 60 |
| O14786 | NRP1_HUMAN | 1 | MERGLPLLCAVLALVLAFAFRNDKCGDTIKIESPGYLTPGYPHSHYHSEKCEWLIQA | 60 |
| Q9QWJ9 | NRP1_RAT | 1 | MERGLPLLCLALALALAGAFRSDKCGGTIKIENPGYLTPGYPHSHYHSEKCEWLIQA | 60 |
| P97333 | NRP1_MOUSE | 61 | PEFYQRIMINFNPHFDLEDRDCKYDYVEVDGENEGGRLWGKFCGKIAPSPVSSGGPFLF | 120 |
| O14786 | NRP1_HUMAN | 61 | PDFYQRIMINFNPHFDLEDRDCKYDYVEVDGENENGFHFKGFCGKIAPSPVSSGGPFLF | 120 |
| Q9QWJ9 | NRP1_RAT | 61 | PEFYQRIMINFNPHFDLEDRDCKYDYVEVDGENEGGRLWGKFCGKIAPSPVSSGGPFLF | 120 |
| P97333 | NRP1_MOUSE | 121 | IKFVSDYETHGAGFSIRYEIIFKRGPECSQNTAPTGVKISPGFPEKYPNSLECTYIIFAP | 180 |
| O14786 | NRP1_HUMAN | 121 | IKFVSDYETHGAGFSIRYEIIFKRGPECSQNTTPSGVIKSPGFPEKYPNSLECTYIIFVFP | 180 |
| Q9QWJ9 | NRP1_RAT | 121 | IKFVSDYETHGAGFSIRYEIIFKRGPECSQNTAPTGVKISPGFPEKYPNSLECTYIIFAP | 180 |
| P97333 | NRP1_MOUSE | 181 | KMSEIILEFESFDLEQDSNPPGGMFCRYDRLEIWDGFFVPGPHIGRYCGQKTPGRIRSSS | 240 |
| O14786 | NRP1_HUMAN | 181 | KMSEIILEFESFDLEPDSNPPGGMFCRYDRLEIWDGFFVPGPHIGRYCGQKTPGRIRSSS | 240 |
| Q9QWJ9 | NRP1_RAT | 181 | KMSEIILEFESFDLEQDSNPPGGVFCRYDRLEIWDGFFVPGPHIGRYCGQKTPGRIRSSS | 240 |
| P97333 | NRP1_MOUSE | 241 | GVLSMVFYTDLSAIAKEGFSANYSVLQSSISEDFKCMALGMESEIHSQDITASSQYGTN | 300 |
| O14786 | NRP1_HUMAN | 241 | GILSMVFYTDLSAIAKEGFSANYSVLQSSVSEDFKCMALGMESEIHSQDITASSQYSTN | 300 |
| Q9QWJ9 | NRP1_RAT | 241 | GILSMVFYTDLSAIAKEGFSANYSVLQSSISEDFKCMALGMESEIHSQDITASSQYGTN | 300 |
| P97333 | NRP1_MOUSE | 301 | WSVERSRLNYPENGWTPGEDSYKEWIQVDLGLLRFVAVGTQGAISKETKKKYVKTYYRV | 360 |
| O14786 | NRP1_HUMAN | 301 | WSAERSRLNYPENGWTPGEDSYREWIQVDLGLLRFVAVGTQGAISKETKKKYVKTYYKI | 360 |
| Q9QWJ9 | NRP1_RAT | 301 | WSVERSRLNYPENGWTPGEDSYREWIQVDLGLLRFVAVGTQGAISKETKKKYVKTYYRV | 360 |
| P97333 | NRP1_MOUSE | 361 | DISSNGEDWITLKEGNKAIIFQGNINPTDVVAVFPEKLIITRFVRIKPAWETGISMRF | 420 |
| O14786 | NRP1_HUMAN | 361 | DVSSNGEDWITLKEGNKPVLFQGNINPTDVVAVFPEKLIITRFVRIKPAWETGISMRF | 420 |
| Q9QWJ9 | NRP1_RAT | 361 | DISSNGEDWITLKEGNKAIIFQGNINPTDVVAVFPEKLIITRFVRIKPAWETGISMRF | 420 |
| P97333 | NRP1_MOUSE | 421 | VYGGKITDYPSCGMLGMVSLISDSQITASNQADRNMWPNIRLVTSRITGVALPPSPHPY | 480 |
| O14786 | NRP1_HUMAN | 421 | VYGGKITDYPSCGMLGMVSLISDSQITSSNQADRNMWPNIRLVTSRITGVALPPSPHPY | 480 |
| Q9QWJ9 | NRP1_RAT | 421 | VYGGKITDYPSCGMLGMVSLISDSQITASNQADRNMWPNIRLVTSRITGVALPPSPHPY | 480 |
| P97333 | NRP1_MOUSE | 481 | TNEWLQVDLGEKIVRGVIIQGGKHRENKVFMRKFKIAYSNNNGSDWKTIMDDSKRKAQSF | 540 |
| O14786 | NRP1_HUMAN | 481 | INEWLQVDLGEKIVRGVIIQGGKHRENKVFMRKFKIAYSNNNGSDWKTIMDDSKRKAQSF | 540 |
| Q9QWJ9 | NRP1_RAT | 481 | TNEWLQVDLGEKIVRGVIIQGGKHRENKVFMRKFKIAYSNNNGSDWKTIMDDSKRKAQSF | 540 |
| P97333 | NRP1_MOUSE | 541 | EGNNNYDTPELRTFPLSTRFIRIYPERATHSGGLRLMELGCEVEAPTAGTTPNGNPF | 600 |
| O14786 | NRP1_HUMAN | 541 | EGNNNYDTPELRTFPALSTRFIRIYPERATHSGGLRLMELGCEVEAPTAGTTPNGNLV | 600 |
| Q9QWJ9 | NRP1_RAT | 541 | EGNNNYDTPELRTFPLSTRFIRIYPERATHSGGLRLMELGCEVEAPTAGTTPNGNPF | 600 |
| P97333 | NRP1_MOUSE | 601 | DECDQDQANCHSGTGGDFQLTGGITVLAIEKPTIIDSTIQSEFFTYGFCNCFGWGSHKTF | 660 |
| O14786 | NRP1_HUMAN | 601 | DECDQDQANCHSGTGGDFQLTGGITVLAIEKPTIIDSTIQSEFFTYGFCNCFGWGSHKTF | 660 |
| Q9QWJ9 | NRP1_RAT | 601 | DECDQDQANCHSGTGGDFQLTGGITVLAIEKPTIIDSTIQSEFFTYGFCNCFGWGSHKTF | 660 |
| P97333 | NRP1_MOUSE | 661 | CHWEHDSHAQLRWSVLTSTKGTPIQDHTGDGNFIYSQADENQKGVARLVSEVYVYQSSAH | 720 |
| O14786 | NRP1_HUMAN | 661 | CHWEHDNHVQLKWSVLTSTKGTPIQDHTGDGNFIYSQADENQKGVARLVSEVYVYQNSAH | 720 |
| Q9QWJ9 | NRP1_RAT | 661 | CHWEHDSHAQLRWRVLTSTKGTPIQDHTGDGNFIYSQADENQKGVARLVSEVYVYQSSAH | 720 |
| P97333 | NRP1_MOUSE | 721 | CMTFWYHMSGSHVGTLRVKLRVYQKPEEYDQLVWVAVGQDHWKEGRVLLHKSILKLYQVI | 780 |
| O14786 | NRP1_HUMAN | 721 | CMTFWYHMSGSHVGTLRVKLRVYQKPEEYDQLVWMAIGHQDHWKEGRVLLHKSILKLYQVI | 780 |
| Q9QWJ9 | NRP1_RAT | 721 | CMTFWYHMSGSHVGTLRVKLRVYQKPEEYDQLVWVAVGQDHWKEGRVLLHKSILKLYQVI | 780 |
| P97333 | NRP1_MOUSE | 781 | FEFEGIGKGNLGGIAVDDISINNHISQEDCAKPTDLKKNTEIKIDETGSTPGYEGEGEGD | 840 |
| O14786 | NRP1_HUMAN | 781 | FEFEGIGKGNLGGIAVDDISINNHISQEDCAKPADLKKNTEIKIDETGSTPGYEGEGEGD | 840 |
| Q9QWJ9 | NRP1_RAT | 781 | FEFEGIGKGNLGGIAVDDISINNHISQEDCAKPTDLKKNTEIKIDETGSTPGYE-EGKGD | 839 |
| P97333 | NRP1_MOUSE | 841 | KNISRKPGNVLKTLDPILITIIAMSALGVLLGAVCGVVLVYACWCHNGMSEKRNLSALENNY | 900 |
| O14786 | NRP1_HUMAN | 841 | KNISRKPGNVLKTLDPILITIIAMSALGVLLGAVCGVVLVYACWCHNGMSEKRNLSALENNY | 900 |
| Q9QWJ9 | NRP1_RAT | 840 | KNISRKPGNVLKTLDPILITIIAMSALGVLLGAVCGVVLVYACWCHNGMSEKRNLSALENNY | 899 |
| P97333 | NRP1_MOUSE | 901 | FELVDGVKLLKDKLNQSNYSEA | 923 |
| O14786 | NRP1_HUMAN | 901 | FELVDGVKLLKDKLNQSTYSEA | 923 |
| Q9QWJ9 | NRP1_RAT | 900 | FELVDGVKLLKDKLNQSNYSEA | 922 |

Performance check:

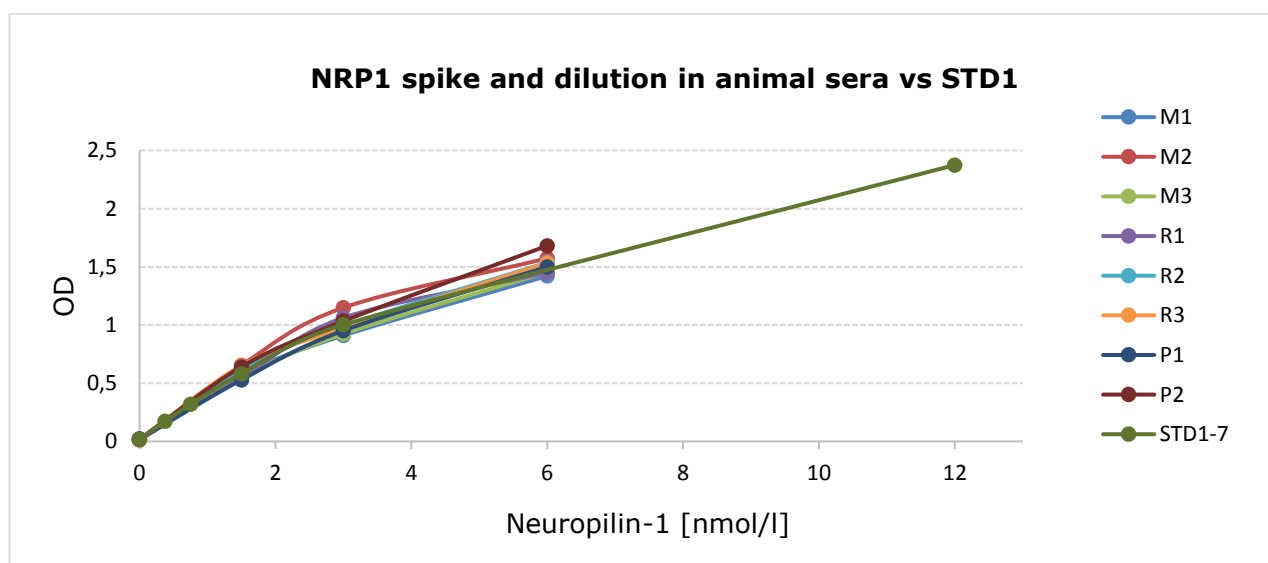
3 mouse serum samples, 3 rat serum samples and 2 pig serum samples (all sera derived from apparently healthy animals) were tested undiluted and spiked by adding STD7 (final concentration 6 nmol/l of human recombinant Neuropilin-1) directly to all samples (ratio 1+1). The spiked sera were diluted 1+1 and 1+3 with STD1 (containing 0 nmol/l NRP1, supplied in the kit). Sample concentrations were calculated by using STD1-STD7.

- Endogenous Neuropilin-1 was undetectable in tested samples thus no test on specificity (competition) was carried out.
- Animal sera can be spiked with recombinant human NRP1 (final concentration 6 nmol/l) – the average recovery is 90%.
- Spiked animal samples can be diluted 1+1 and 1+3 with STD1 (0 nmol/l) – the average recovery is 111% and 119%.

Table: calculation of animal sample concentrations, and spike recovery

| Sample matrix | ID | Ref | NRP1 +6 nmol/l | S/R [%] | dil 1+1 | dil 1+3 | R [%] 1+1 | R [%] 1+3 | |
|---------------|----|-----|----------------|-------------------|-----------|---------|-----------|------------|------------|
| mouse serum | M1 | 0 | 4.9 | 81 | 2.6 | 1.5 | 108 | 121 | |
| mouse serum | M2 | 0 | 5.7 | 95 | 3.6 | 1.7 | 126 | 123 | |
| mouse serum | M3 | 0 | 5.0 | 84 | 2.7 | 1.4 | 107 | 112 | |
| rat serum | R1 | 0 | 5.0 | 84 | 3.2 | 1.4 | 129 | 112 | |
| rat serum | R2 | 0 | 5.5 | 92 | 2.9 | 1.6 | 107 | 119 | |
| rat serum | R3 | 0 | 5.5 | 91 | 2.9 | 1.7 | 105 | 127 | |
| pig serum | P1 | 0 | 5.3 | 88 | 2.8 | 1.4 | 106 | 102 | |
| pig serum | P2 | 0 | 6.3 | 106 | 3.1 | 1.7 | 98 | 107 | |
| | | | | Mean R [%] | 90 | | | 111 | 119 |

Graph: Neuropilin-1 standard curve (STD1-7) and dilution curves of animal sera spiked with human NRP1 diluted with STD1 (0 nmol/l NRP1)



Suggested protocol for the measurement of human Neuropilin-1 in non-human samples

Follow standard protocol as indicated in the package insert:

In pre-dilution plate:

*Use **10 µl non-human sample for GuHCl treatment.***

Continue with protocol as indicated in the instructions for use.

In pre-coated plate:

*Transfer **50 µl pre-treated non-human sample** from pre-dilution plate into respective wells. Swirl gently.*

For the transfer into the coated plate it is recommended to use a multichannel pipette. Transfer should be performed as soon as possible.

Continue with protocol as indicated in the instructions for use.

If required, dilute samples 1+1 with STD1 (provided in the kit) prior to GuHCl treatment.